

An Index of Protein Hydrophobicity. Its Application to Membrane Proteins

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A hydrophobicity index is proposed for proteins. The calculation of this index is made, assuming a direct relationship between the hydrophobicity of each aminoacid and its Rf in a partition chromatography. This index is applied to membrane proteins and offers statistically significant differences between integral and peripheral proteins.

Biological membrane proteins may be divided into two groups, namely *peripheral* and *integral* proteins (26). Integral proteins are strongly and, perhaps, specifically bound to membrane lipids, whereas peripheral proteins are more loosely bound to the membrane core.

It is not known what makes a protein to behave as integral or peripheral, or else as a *soluble* protein, unrelated to membranes. It has been proposed that integral proteins would possess hydrophobic and hydrophilic peptides in contact with the lipid and aqueous phase respectively.

Aminoacid sequence determination of cytochrome b₅ has actually shown the existence of these hydrophilic and hydrophobic peptides (21, 31). It is also possible that, although these two types of zones would not be identified by the study of the primary structure, a given protein could show an amphipathic character because of its tertiary structure.

Attempts have been made (2, 7) to relate the more or less hydrophobic character of a protein, and therefore its integral or peripheral nature, to its aminoacid composition. These attempts have not been proved completely satisfactory. In this report an index for measuring protein hydrophobicity is proposed, assuming a direct relationship between the hydrophobicity of each aminoacid and its Rf in a partition chromatography. Statistically significant differences can be observed

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between hydrophobicity indexes (H.I.) of integral and peripheral proteins.

Materials and Methods

Calculation of the H.I. This index is based on the assumption that the hydrophobicity of a given protein may be estimated by adding the hydrophobicities due to each aminoacid. The hydrophobicities of the different aminoacids are assimilated to their respective $Rf \times 100$ values in a chromatographic system based mainly on partition processes.

The $Rf \times 100$ values chosen have been those cited by SMITH (27) for paper chromatography using butanol-pyridine-water (1:1:1) as solvent. These values are reproduced in table I. The H.I. is computed from the percent composition in aminoacids of the protein. Percent molar values of each aminoacid are multiplied by the

respective $Rf's \times 100$, and added. The total sum is the so-called «hydrophobicity index» (H.I.).

Aminoacid composition of ten integral, ten peripheral and ten soluble proteins was taken from the literature. The validity of our results was tested by means of a Model 1 single classification analysis of the variance, according to SOKAL and ROHLF (28).

Results and Discussion

Table II shows the H.I. of the thirty proteins mentioned above.

The analysis of the variance was significant ($p < 0.01$). Thus, comparisons among means were established using an *a posteriori* SS-STP test (28).

Integral proteins had an H.I. significantly higher than peripheral and soluble proteins. No significant difference was found between peripheral and soluble proteins.

The H.I. was also calculated for the different subunits of proteins showing complex quaternary structure, such as the oligomycin-sensitive mitochondrial ATPase. This enzyme has a spherical «head», which protrudes out of the membrane into the water phase, and is linked through a «stalk» to a «base piece». Interestingly enough, the «head» polypeptides (24) have an average value for H.I. of 3266, which accounts for a protein of extrinsic nature, while one of the main polypeptides of the base piece, described by TZAGOLOFF *et al.* (34), which is supposed to be deeply buried into the lipid core, has the extremely high H.I. value of 4109, which corresponds to a protein of a highly integral character.

The validity of the H.I. was tested by calculating the corresponding values for the 30 proteins on different data: namely, Rf values cited by SMITH (27) for paper chromatography using butanol-acetic acid-water (12:3:5) as solvent. This mobile phase is acid, in contrast with butanol-

Table I. $Rf \times 100$ values of the 20 common aminoacids

These Rf values correspond to an ascending partition chromatography, using butanol-pyridine-water (1:1:1) as solvent (27).

Name	$Rf \times 100$
Lysine	13
Cysteine	14
Arginine	15
Aspartic acid	20
Glutamic acid	20
Histidine	24
Glycine	29
Hidroxypoline	30
Serine	33
Proline	34
Threonine	36
Alanine	37
Valine	48
Methionine	53
Isoleucine	56
Leucine	60
Tyrosine	60
Tryptophan	62
Phenylalanine	63

Table II. *Hydrophobicity indexes of ten integral, ten peripheral and ten soluble proteins.*
For details see text.

	H.I.	Ref.		H.I.	Ref.			
<i>Integral proteins</i>								
C _{ss} Isoprenyl alcohol phosphokinase	4261	22	Monoamine oxidase, mitochondrial	3530	20			
Carotenoid glycoprotein, <i>Sarcina flava</i>	4070	33	Calsequestrin, sarcoplasmic reticulum	3365	12			
Folch-Lees proteolipid protein, sciatic myelin	4064	6	Cytochrome c, human	3297	16			
Cytochrome b, mitochondrial	4019	19	Basic protein, sciatic myelin	3203	6			
Proteolipid, sarcoplasmic reticulum	3936	13	Basic protein, brain myelin	3154	6			
Cytochrome a, mitochondrial beef heart	3880	15	54 K protein, sarcoplasmic reticulum	3107	13			
Purple membrane protein of <i>Halobacterium halobium</i>	3878	30	Mean H.I. value±S.D.	3396±91				
Rhodopsin, bovine	3791	25	<i>Soluble proteins</i> (unrelated to membranes)					
Sarcoplasmic reticulum ATPase	3611	11	Acetoacetate decarboxilase	3662	10			
Cytochrome b _s , microsomal	3503	29	Alanine aminotransferase	3599	17			
Mean H.I. value±S.D.	3901±71		Ceruloplasmin	3598	9			
<i>Peripheral proteins</i>			Adenylate kinase	3595	14			
D-glyceraldehyde-3-phosphate dehydrogenase	3601	32	Fumarate hydratase	3579	8			
ATPase, <i>Bacillus megaterium</i>	3591	18	Molate dehydrogenase	3546	5			
ATPase, mitochondrial	3567	3	Phosphoenolpyruvate carboxykinase	3530	4			
ATPase, <i>Streptococcus faecalis</i>	3544	23	D-Amylase	3524	35			
			K-Casein (component A-1)	3480	36			
			Chorionic gonadotropin	3288	1			
			Mean H.I. value±S.D.	3540±32				

pyridine-water, which is alkaline, and thus the relative mobilities of the ionizable aminoacids are different. However, similar significant differences were found between the indexes of peripheral and integral proteins.

These results suggest that a direct relationship exists between the aminoacid composition of a membrane protein and its relation to membrane structure. The hydrophobicity index permits in this way

to predict with reasonable accuracy the integral or peripheral character of a protein whose aminoacid composition has been investigated after polyacrylamide gel electrophoresis of a complex mixture.

Resumen

Se propone un índice de hidrofobicidad aplicable a las proteínas. El cálculo de este índice se realiza suponiendo que existe una relación

directa entre el índice de hidrofobicidad de cada aminoácido y su Rf en una cromatografía de partición. Al aplicar este índice a proteínas de membranas se observa la existencia de diferencias estadísticamente significativas entre proteínas integrales y periféricas.

References

1. BAHL, O. P.: *J. Biol. Chem.*, **244**, 567-576, 1969.
2. CAPALDI, R. and VANDERKOOI, G.: *Proc. Nat. Acad. Sci. U.S.A.*, **69**, 930-933, 1972.
3. CATTERALL, W. A. and PEDERSEN, P. L.: *J. Biol. Chem.*, **241**, 2413-2420, 1971.
4. CHANG, H. C. and LANE, M. D.: *J. Biol. Chem.*, **241**, 2413-2420, 1966.
5. CONSIGLIO, E., VARRONE, S. and COVELLI, I.: *Eur. J. Biochem.*, **17**, 408-414, 1970.
6. ENG, L. P., CHAO, F. C., GERSTL, B., PRATT, D. and TASTSTJEMA, M. G.: *Biochemistry*, **7**, 4455-4465, 1968.
7. EPSTEIN, C. J.: *Nature*, **215**, 355-359, 1967.
8. KANAREK, L. and HILL, R. L.: *J. Biol. Chem.*, **239**, 4202-4206, 1964.
9. KASPER, C. B. and DEUTSCH, M. F.: *J. Biol. Chem.*, **238**, 2325-2337, 1963.
10. LEDERER, F., COULTS, S. M., LAURSEN, R. A. and WESTHEIMER, F. M.: *Biochemistry*, **5**, 823-833, 1966.
11. MACLENNAN, D. H., SEEMAN, P., ILES, M. and YIP, C. C.: *J. Biol. Chem.*, **246**, 2702-2710, 1971.
12. MACLENNAN, D. H. and WONG, P. T. S.: *Proc. Nat. Acad. Sci. U.S.A.*, **68**, 1231-1235, 1971.
13. MACLENNAN, D. H., YIP, C. C., ILES, G. M. and SEEMAN, P.: In «Cold Spring Harbor Symposia on Quantitative Biology», Vol. 37, pp. 469-477, 1972.
14. MARKLAND, F. E. and WADKINS, C. L.: *J. Biol. Chem.*, **241**, 4136-4145, 1966.
15. MATSUBARA, T., ORII, Y. and OKURUKI, K.: *Biochem. Biophys. Acta*, **97**, 61-67, 1965.
16. MITSUBARA, H. and SMITH, E. L.: *J. Biol. Chem.*, **237**, 3575-3576, 1962.
17. MATSUZAWA, T. and LEGAL, M. L.: *J. Biol. Chem.*, **243**, 5929-5934, 1968.
18. MIRSKY, R. and BARLOW, V.: *Biochem. Biophys. Acta*, **291**, 480-492, 1973.
19. OHNISHI, S. and ITO, T.: *Biochem. Biophys. Res. Comm.*, **51**, 132-138, 1973.
20. ORELAND, L.: *Arch. Biochem. Biophys.*, **146**, 410-424, 1971.
21. OZOLS, J. and STRITTMATTER, P.: *J. Biol. Chem.*, **244**, 6617-6624, 1969.
22. SANDERMANN, H., Jr., and STROMINGER, J. L.: *Proc. Nat. Acad. Sci. U.S.A.*, **68**, 2441-2445, 1971.
23. SCHENEBLE, H. P., VATTER, A. E. and ABRAMS, A.: *J. Biol. Chem.*, **245**, 1122-1127, 1970.
24. SENIOR, A. E. and MACLENNAN, D. H.: *J. Biol. Chem.*, **245**, 5086-5095, 1970.
25. SHIELDS, J. E., DINOV, E., HENDRICKSEN, R., KIMBEL, R. and MILLAR, P.: *Biochem. Biophys. Acta*, **147**, 238-251, 1969.
26. SINGER, S. J.: *Ann. Rev. Biochem.*, **45**, 805-833, 1974.
27. SMITH, I.: In «Chromatographic and Electrophoretic Techniques» (Smith, I., ed.), Vol. I, pp. 75-108, W. Heinemann, London, 1976.
28. SOKAL, R. R. and ROHLF, J.: «Introduction to Biostatistics», W. M. Freeman and Co., San Francisco, 1969.
29. SPATZ, L. and STRITTMATTER, P.: *J. Biol. Chem.*, **248**, 793-799, 1973.
30. STOECKENIUS, W. and KUNAN, W. H.: *J. Cell Biology*, **38**, 337-357, 1968.
31. STRITTMATTER, P., ROGERS, M. J. and SPATZ, L.: *J. Biol. Chem.*, **247**, 7188-7196, 1972.
32. TANNER, M. J. A. and GRAY, W. R.: *Biochem. J.*, **125**, 1109-1117, 1971.
33. THURKELL, O. and HUNTER, M. S. I.: *J. Gen. Microbiol.*, **58**, 289-292, 1969.
34. TZAGOLOFF, A., AKAI, A., RUBIN, M. S.: In «The Biogenesis of Mitochondria» (Kroon, A. M. and Saccone, C., eds.), Academic Press, New York and London, 1973, pp. 405-421.
35. VANDERMEERS, A. and CHRISTOPE, J.: *Biochem. Biophys. Acta*, **154**, 110-129, 1968.
36. WOYCHIK, J. M., KALAN, E. B. and NOEKEN, M. E.: *Biochemistry*, **5**, 2276-2282, 1966.